

Product Information

SYBR Safe DNA Gel Stain, 10,000X

Catalog Number	Packaging Size
N107	400 µL

Storage upon receipt:

- 2-25°C
- Protect from light

Ex/Em: 500/530 nm, bound to nucleic acid

Product Description

SYBR Safe stain is a stable and environmentally safe green fluorescent nucleic acid dye specifically designed for gel staining. **SYBR Safe stain** is compatible with a standard 300 nm transilluminator, a 254 nm transilluminator, a blue-light transilluminator, or a gel reader equipped with visible light excitation such as a 488 nm laser-based gel scanner.

SYBR Safe DNA Gel Stain, 10,000X is a concentrated **SYBR Safe stain** solution that can be diluted 10,000 times for use in precast gel staining or 5,000 times for use in post gel staining according to the procedures described below. One vial of 10,000X solution can be used to prepare at least 100 precast minigels or post-stain at least 100 minigels.

Gel staining with **SYBR Safe stain** is compatible with downstream applications such as gel extraction and cloning. **SYBR Safe stain** is efficiently removed from DNA by phenol/chloroform extraction and ethanol precipitation.

Staining Protocols

1. Post-staining Protocol

- 1.1 Run gels as usual according to your standard protocol.
- 1.2 Dilute the **SYBR Safe stain 10,000X** stock reagent 5,000 fold to make a 2X staining solution in TE, TBE, or TAE buffer.
- 1.3 Carefully place the gel in a suitable polypropylene container. Gently add a sufficient amount of the 2X staining solution to submerge the gel.

- 1.4 Agitate the gel gently at room temperature for 30 min.

- 1.5 Wash the gel with DI water to remove excess dye. Image the stained gel with a transilluminator, or a laser-based gel scanner using a long path green filter such as a SYBR Filter or GelStar filter.

2. Pre-cast Protocol

- 2.1 Prepare molten agarose gel solution using your standard protocol.

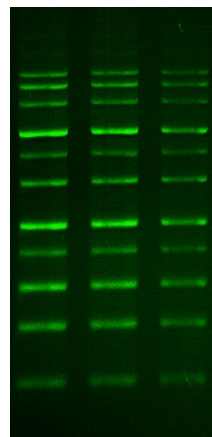
- 2.2 Dilute the **SYBR Safe stain 10,000X** stock reagent into the molten agarose gel solution at 1:10,000 and mix thoroughly.

- 2.3 Cast the gel and allow it to solidify.

- 2.4 Load samples and run the gels using your standard protocol.

- 2.5 Image the stained gel with a transilluminator, or a laser-based gel scanner using a long path green filter such as a SYBR Filter or GelStar filter.

Note: The pre-cast protocol is not recommended for polyacrylamide gels. Use the post staining protocol for acrylamide gels.



SYBR Safe stain in pre-cast gel staining