

Product Information

Mitochondrion-Selective Probes

Catalog Number	Product Name	Unit Size
C040	MitoTrack Green FM	1 mg
C041	MitoTrack Orange CMTMRos	1 mg
C042	MitoTrack Red CMXRos	1 mg
C054	MitoTrack Red FM	1 mg
C043	MitoTrack Deep Red FM	1 mg

Storage upon receipt:

- -20°C
- Protect from light

Product Description

The cell-permeant MitoTrack probes contain a mildly thiol-reactive chloromethyl moiety for labeling mitochondria, and are retained in the mitochondria after fixation.

To label mitochondria, cells are simply incubated with MitoTrack probes, which passively diffuse across the plasma membrane and accumulate in active mitochondria. Once their mitochondria are labeled, the cells can be treated with an aldehyde-based fixative for samples that need fixation to allow further processing of the sample. These MitoTrack probes eliminate some of the difficulties of working with pathogenic cells because once the mitochondria are stained, the cells can be treated with fixatives before the sample is analyzed.

Although conventional fluorescent stains for mitochondria, such as tetramethylrhodamine and rhodamine 123, are readily sequestered by functioning mitochondria, these stains are easily washed out of cells once the mitochondria experience a loss in membrane potential. This characteristic limits the use of such conventional stains in experiments that require cells to be treated with aldehyde fixatives or with other agents that affect the energetic state of the mitochondria. To overcome this limitation, use our MitoTrack probes that are concentrated by active mitochondria and retained during cell fixation.

Table. Spectral characteristics of the MitoTrack probes

Product Name	Ex	Em
MitoTrack Green FM	490 nm	516 nm
MitoTrack Orange CMTMRos	554 nm	576 nm
MitoTrack Red CMXRos	579 nm	600 nm
MitoTrack Red FM	580 nm	644 nm
MitoTrack Deep Red FM	645 nm	665 nm

Experimental Protocols

Cell Preparation and Staining

The concentration of probe for optimal staining varies by application. The initial conditions suggested here are guidelines that may be modified based on the particular cell type or on other factors, such as the permeability of the cells or tissues to the probe.

1.1 Preparing staining solutions. Dissolve the MitoTrack probe in anhydrous DMSO to a final concentration of 2 mM. Dilute 2 mM MitoTrack stock solution to the final working concentration in appropriate buffer or growth medium.

General Guidelines: Use working concentrations of 25-500 nM. For staining cells that are to be fixed and permeabilized (see *Fixation and Permeabilization after Staining*), use a working concentration of 100-500 nM. To reduce potential artifacts and mitochondrial toxicity from overloading, keep the concentration of dye as low as possible. For the MitoTrack Green FM probe, use a slightly lower concentration (20-200 nM). At higher concentrations, these probes tend to stain other cellular structures.

1.2 Staining adherent cells. Grow cells on coverslips inside a Petri dish filled with the appropriate culture medium. When cells have reached the desired confluency, remove the media from the dish and add prewarmed (37°C) staining solution containing MitoTrack probe (prepared in step 1.1). While incubation times vary depending on the model system and probe used, incubation for 15-45 minutes under growth conditions appropriate for the particular cell type is generally sufficient but may need to be optimized. After staining is complete replace the staining solution with fresh prewarmed media or buffer and observe cells using a fluorescence microscope or fluorescence microplate reader. If the cells are to be fixed and permeabilized, continue to *Fixation and Permeabilization after Staining*.

1.3 Staining suspension cells. Centrifuge to obtain a cell pellet and aspirate the supernatant. Resuspend the cells gently in prewarmed (37°C) staining solution containing the MitoTrack probe (prepared in step 1.1). While incubation times vary depending on the model system and probe used, incubation for 15-45 minutes under growth conditions appropriate for the particular cell type is generally sufficient but may need to be optimized.

After staining is complete, re-pellet the cells by centrifugation and resuspend cells in fresh prewarmed medium or buffer. Cells may be analyzed by flow cytometry, microplate-based analysis, or fluorescence microscopy. If immobilized cells on

coverslips are needed, use poly-D-lysine to coat the slides or coverslips before mounting. If the cells are to be fixed and permeabilized, continue to *Fixation and Permeabilization after Staining*.

Optional: Fixation and Permeabilization after Staining

After staining live cells with a MitoTrack dye, it is often useful to fix and permeabilize the cells for subsequent manipulations. For example, fixation and permeabilization allows you to probe for other intracellular structures by immunocytochemistry.

2.1 Washing the cells. After staining, wash the cells in fresh, pre-warmed buffer or growth medium.

2.2 Fixing the cells. Carefully remove the medium/buffer covering the cells, and replace it with freshly prepared, pre-warmed buffer or growth medium containing 2-4% formaldehyde.

2.3 Rinsing the cells. After fixation, rinse the cells several times in buffer.

2.4 Permeabilization (optional). When permeabilization is needed for subsequent steps such as immunocytochemistry, incubate fixed cells in buffer containing detergent such as Triton X-100. Following permeabilization, rinse the cells in buffer and proceed with immunocytochemistry procedure. We found that incubating the endothelial cells for 10 minutes in PBS containing 0.2% Triton X-100 works well.

Alternatively, the cells may be permeabilized by incubating in ice-cold acetone for 5 minutes, and then washed in PBS.