



## Safety Report

### Product and Company Identification

Product Name	RedView DNA Gel Stain
Product Number	N102
Unit Size	500 µL
Manufacturer/Supplier	ABP Biosciences 7040 Virginia Manor Road Beltsville, MD 20705, USA Web: <a href="http://www.abpbio.com">www.abpbio.com</a>

### Introduction

Ethidium bromide (EB) is the most commonly used nucleic acid stain in molecular biology laboratories for decades. The dye is inexpensive, sensitive and very stable. However, it has been proved to be strong carcinogen and therefore considered hazardous for laboratory personnel and environment.

To overcome the toxicity problem of EB, ABP Biosciences has developed a RedView DNA Gel Stain as a safer alternative to the traditional EB for detecting nucleic acid in agarose gel. It is as sensitive as EB and can be used exactly the same way in agarose gel electrophoresis with significantly improved safety profiles over EB.

The safety of RedView DNA Gel Stain has been performed with the following four tests:

1. Ames test;
2. Cell membrane permeability and cytotoxicity test;
3. Glove penetration test;
4. Acute oral toxicity test.

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## AMES TEST

### Purpose

The Ames test is a standard assay to assess the mutagenic potential of chemicals. As cancer is often associated with DNA damage, the test can be used to estimate the carcinogenic potential of a chemical compound.

### Test System

The Ames test employed four Salmonella strains, TA97, TA98, TA100 and TA102. When these bacteria are exposed to mutagenic agents, under certain conditions reverse mutation from amino acid (histidine) auxotrophy to prototrophy occurs, giving colonies of revertants. In order to test the mutagenic toxicity of metabolised products, S9 fraction, a rat liver extract, was used in the assays. The S9 fraction contains a mixture of several enzymes and is known to be able to convert some chemicals into mutagens.

### Test Article

RedView DNA Stain was dissolved in the sterile distilled water, and the concentrations were 0 (control), 5, 10, 25, 50, 100, 250 and 500 µg/mL, respectively. The test volume was 0.1 mL per plate.

The control groups included blank control plates, and positive control plates. In the absence of S9 mixture, the positive control reference for strains TA97 and TA98 was 9-fluorenone, for TA100 was  $\text{NaN}_3$ , and for TA102 was Mitomycin C. In the presence of the S9 mixture, the positive control reference substance for strains TA97, TA98 and TA100 was 2-AF (Aminofluorene), and 1,8-hydroxyanthraquinone for TA102.

### Test Procedure

The test substance or control chemical (0.1 mL) and 0.1 mL bacterial suspension with 0.5 mL S9 mixture (+S9) or phosphate buffered saline (-S9) were mixed uniformly in test tubes with 1.5 mL overlay agar (liquid, 45°C). The mixture was uniformly poured on the prepared underlay agar plates. After solidification, the plates were incubated at 37°C for 48 h. At the end of the incubation, revertant colonies per plate were counted. All plating was done in triplicate. If the number was more than twice the spontaneous revertant colonies counts and showed a dose-response relationship, positive result for mutagenicity could be concluded.

### Test Result

According to the results of the Ames test (Table 1), in the presence and absence of metabolic activator S9 the increase in the numbers of revertant colonies of strains TA97, TA98, TA100 and TA102 compared to spontaneous revertant colonies was less than 2 times, and there was no dose-response relationship. Appropriate reference mutagens were used as positive controls and they showed a distinct increase of induced revertant colonies (Table 1).

**Table 1.** Results of RedView DNA Stain Ames test

Dosage μg/plate	TA <sub>97</sub>		TA <sub>98</sub>		TA <sub>100</sub>		TA <sub>102</sub>	
	without S <sub>9</sub>	with S <sub>9</sub>	without S <sub>9</sub>	with S <sub>9</sub>	without S <sub>9</sub>	with S <sub>9</sub>	without S <sub>9</sub>	with S <sub>9</sub>
0	120	122	38	37	105	106	215	218
0.5	121	123	38	40	102	105	213	220
1	123	121	42	44	106	108	216	224
2.5	126	123	40	46	103	110	211	236
5	132	128	42	41	108	106	224	227
10	125	134	39	48	110	121	232	242
25	134	143	41	50	104	126	226	234
50	135	157	44	47	113	116	232	243
Pos. control	1643 <sup>1</sup>	1725 <sup>2</sup>	1489 <sup>1</sup>	1816 <sup>2</sup>	1328 <sup>3</sup>	1223 <sup>2</sup>	1425 <sup>4</sup>	1506 <sup>5</sup>

**Note:** 1. 9-Fluorenone, 0.2 μg/plate;  
2. 2-AF, 10 μg/plate;  
3. NaN<sub>3</sub>, 2.5 μg/plate;  
4. Mitomycin C, 4 μg/plate;  
5. 1,8-Hydroxyanthraquinone, 50 μg/plate.

## Conclusion

RedView DNA Gel Stain is nonmutagenic over the dose range from 0.5 μg/plate (or 225 ng/mL) to 50 μg/plate (or 22.5 μg/mL) to these *Salmonella typhimurium* strains. RedView working concentration used in gel staining is 1-5 μg/mL, which is well within the safety range.

## CELL MEMBRANE PERMEABILITY AND CYTOTOXICITY TEST

### Purpose

The purpose of this test is to see if RedView DNA Gel Stain can cross cell membranes to stain nuclear DNA, and how it will affect the cytotoxicity.

### Method

#### Cell membrane permeability test

Hela cells were cultured in 24-well tissue culture plate. Each well was incubated with SYBR Safe, GreenView, GreenView Plus, and RedView, respectively. The dye concentrations were all 1X based on the respective dye concentrations used for gel staining for each dye. The SYBR Safe was used as a control as it is known to be able to stain DNA in live cells. After incubation at 37 °C for 30 min, cell imaging was taken by fluorescence microscopy using optical filter sets appropriate for each dye.

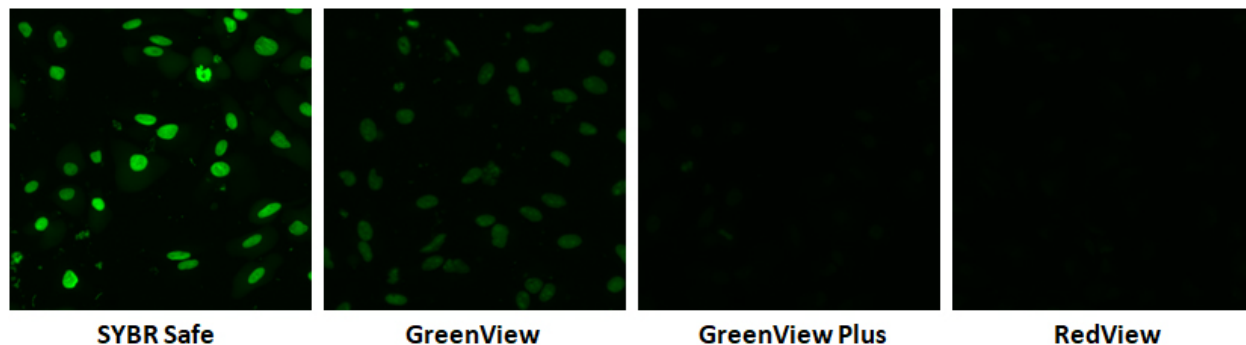
#### Cytotoxicity test

Hela cells were cultured in 96-well tissue culture plate. Each well was incubated with SYBR Safe, GreenView, GreenView Plus, and RedView, respectively. The dye concentrations were 4, 10, 20, 40 µM for each dye. As negative control the cells were treated only with medium. The positive control group was treated with phenol. After incubation at 37 °C for 24 hours, MTT was added in all wells and incubated for 2 hours. After incubation, DMSO was added to the wells and read at 570 nm using a spectrophotometer. Mean value of growth inhibition was calculated by the following formula:

$$\text{Mean value growth inhibition} = 100\% \times [A570_{(\text{Negative control})} - A570_{(\text{test compound})}] / A570_{(\text{Negative control})}$$

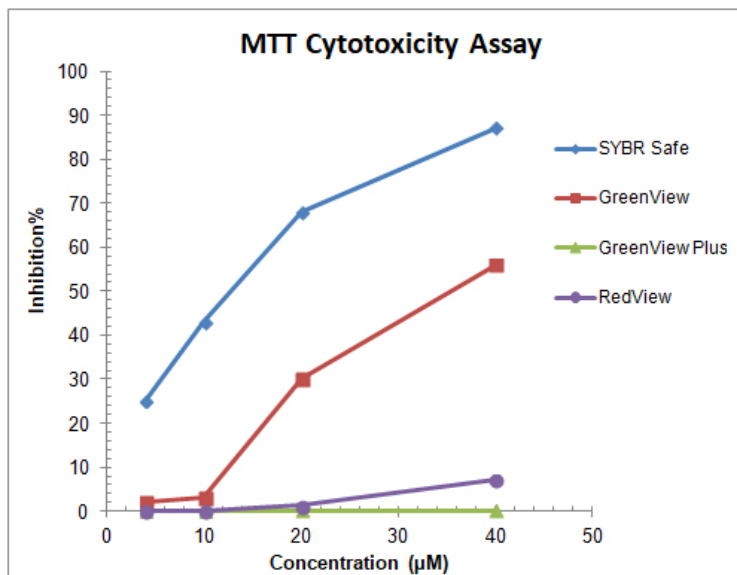
### Results

Cell images obtained following 30 minutes of incubation are shown in Figure 1. Cells stained with SYBR Safe show bright nuclear stain, cells stained with GreenView show weak nuclear stain, and cells stained with GreenView Plus or RedView show no nuclear counterstain.



**Figure 1.** HeLa cells were incubated with 1X of SYBR Safe, GreenView, GreenView Plus, and RedView, respectively. After incubation at 37 °C for 30 min, cell imaging was taken by fluorescence microscopy, respectively.

Results of cytotoxicity assay are shown in Figure 2. SYBR Safe shows moderate inhibition at low concentration at 4  $\mu\text{M}$ . When the concentration of SYBR Safe is increased to 20  $\mu\text{M}$ , the inhibition becomes significantly. When the concentration of GreenView is below 10  $\mu\text{M}$ , there is no inhibition to cells. But when the concentration of GreenView is increased to 20  $\mu\text{M}$ , GreenView will show moderate inhibition. Instead, even the concentration of GreenView Plus or RedView is over 40  $\mu\text{M}$ , there is no or little inhibition to cells. The dye working concentration used in gel staining is about 2-5  $\mu\text{M}$ .



**Figure 2.** MTT cytotoxicity assay with SYBR Safe, GreenView, GreenView Plus, and RedView.

## Conclusion

RedView DNA Gel Stain is impermeable to cell membranes, and has no cytotoxic effects at working concentration.

## GLOVE PENETRATION TEST

### Purpose

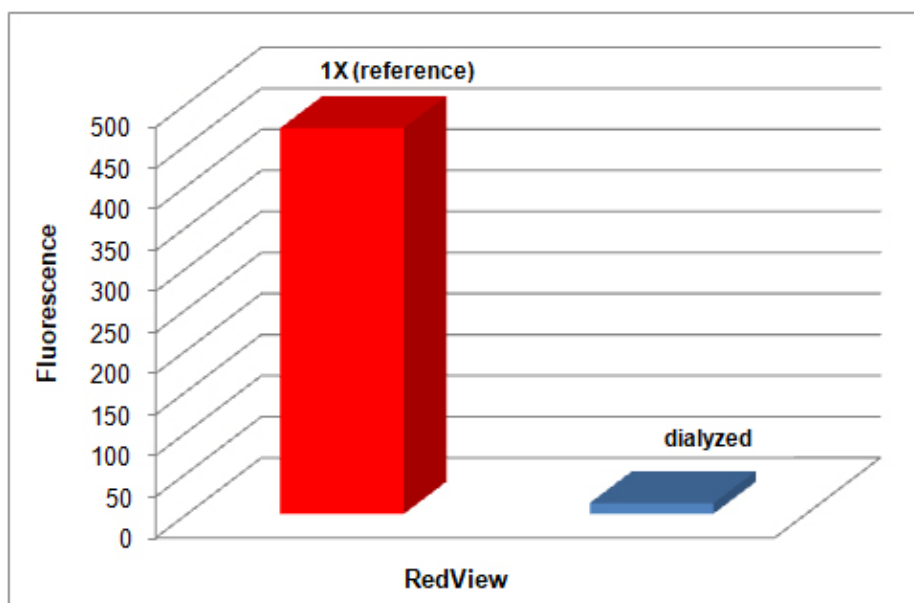
The purpose of the glove penetration tests is to test the ability of RedView DNA Gel stain to diffuse through latex gloves.

### Method

A finger of a latex glove containing 0.5X TBE buffer was dialyzed against 0.5X TBE buffer containing 5X RedView DNA Gel stain for 24 hours. The solution in the finger was then analyzed for presence of the dye by fluorescence. As a reference, the fluorescence of the dye at 1X was also measured. To increase the sensitivity of the detection, all fluorescence measurements were made in the presence of 50 µg/mL salmon sperm dsDNA. Maximum excitation and emission wavelengths were chosen for this analysis (540 nm for excitation and 600 nm for emission). The bandwidths of excitation and emission filters are 10 nm.

### Results

The result of the test shows RedView is impenetrable to latex gloves (Figure 3).



**Figure 3.** Relative fluorescence of solutions dialyzed in latex glove fingers against 5X RedView and the relative fluorescence of the corresponding 1X dye solution as a reference.

### Conclusion

Handling RedView with latex gloves is safe.

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## ACUTE ORAL TOXICITY TEST

### Purpose

The acute toxicity testing provides information on the biologic activity of the test item that can be used in hazard identification and risk management in the context of production, handling, and use of chemicals. This procedure is designed to determine the acute oral toxicity of the material under test.

### Method

1X RedView DNA Gel Stain in 0.5X TBE buffer was directly used as test article. A single dosage at 10g/kg was chosen. The animals used in the test were healthy Kuming mice. Before testing, the animals were fasted for 18 hours but water was ad libitum. Food was withheld until 4 hours after dosing in order to facilitate gastrointestinal absorption of the test article. The animals were administrated with 10 g/kg of the test article solution by means of a gavage needle attached to a hypodermic syringe.

After exposure, the animals were observed for mortality, weight change, and toxic signs for a two-week period.

### Results

All animals remained healthy with no toxic signs throughout the duration of the study. All animals gained weight during the test period. No abnormalities were observed on all animals at the end of the study.

### Conclusion

A single oral administration of RedView DNA Gel Stain in 0.5X TBE at a limit dose of 10 g/kg to three healthy mice produced no mortalities or toxic signs.